

# WEST Search History

DATE: Wednesday, December 11, 2002

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L3	l1 and L2	6	L3
L2	vector same (P1 replicon and factor)	7	L2
L1	method near3 clon\$	12229	L1

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NEWS 7 Apr 22 BIOSIS Gene Names now available in TOXCENTER  
NEWS 8 Apr 22 Federal Research in Progress (FEDRIP) now available  
NEWS 9 Jun 03 New e-mail delivery for search results now available  
NEWS 10 Jun 10 MEDLINE Reload  
NEWS 11 Jun 10 PCTFULL has been reloaded  
NEWS 12 Jul 02 FOREGE no longer contains STANDARDS file segment  
NEWS 13 Jul 22 USAN to be reloaded July 28, 2002;  
saved answer sets no longer valid  
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NEWS 15 Jul 30 NETFIRST to be removed from STN  
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NEWS 17 Aug 08 PHARMAMarketLetter(PHARMAML) - new on STN  
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NEWS 22 Aug 26 Sequence searching in REGISTRY enhanced  
NEWS 23 Sep 03 JAPIO has been reloaded and enhanced  
NEWS 24 Sep 16 Experimental properties added to the REGISTRY file  
NEWS 25 Sep 16 Indexing added to some pre-1967 records in CA/CAPLUS  
NEWS 26 Sep 16 CA Section Thesaurus available in CAPLUS and CA  
NEWS 27 Oct 01 CASREACT Enriched with Reactions from 1907 to 1985  
NEWS 28 Oct 21 EVENTLINE has been reloaded  
NEWS 29 Oct 24 BEILSTEIN adds new search fields  
NEWS 30 Oct 24 Nutraceuticals International (NUTRACEUT) now available on STN  
NEWS 31 Oct 25 MEDLINE SDI run of October 8, 2002  
NEWS 32 Nov 18 DKILIT has been renamed APOLLIT  
NEWS 33 Nov 25 More calculated properties added to REGISTRY  
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AND CURRENT DISCOVER FILE IS DATED 01 OCTOBER 2002  
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=> s vector (3s) (p1 replicon and factor)  
PROXIMITY OPERATION NOT ALLOWED  
PROXIMITY OPERATION NOT ALLOWED  
PROXIMITY OPERATION NOT ALLOWED  
Certain operators may not be nested in combination with other operators. A nested operator is valid only when it occurs at the same level or above the operator outside the nested phrase as determined by the following precedence list:

1. Numeric
2. (W), (NOTW), (A), (NOTA)
3. (S), (NOTS)
4. (P), (NOTP)
5. (L), (NOTL)
6. AND, NOT
7. OR

For example, '(MONOCLONAL(W)ANTIBOD?)(L)ANTIGEN?' is valid since (W) is above (L) on the precedence list. However, '((THIN(W)LAYER)(L)PHOSPHOLIPID#)(A)LACTONE#' is not valid since (L) is below (A) on the precedence list. The only exception is the 'OR' operator. This operator may be used in combination with any other operator. For example, '(ATOMIC OR NUCLEAR)(W)REACTOR' is valid.

=> s p1 replicon and F factor  
L1 1 P1 REPLICON AND F FACTOR

=> d bib abs

L1 ANSWER 1 OF 1 CAPLUS COPYRIGHT 2002 ACS  
AN 1999:90263 CAPLUS  
DN 130:134980  
TI Shuttle vectors containing replication origins and selection markers for both yeast and bacteria  
IN Bradshaw, M. Suzanne; Bollekens, Jacques A.; Ruddle, Frank H.  
PA Yale University, USA  
SO U.S., 15 pp.  
CODEN: USXXAM  
DT Patent  
LA English  
FAN.CNT 2

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI US 5866404	A	19990202	US 1996-761704	19961206
US 5972614	A	19991026	US 1997-987966	19971210
US 6221588	B1	20010424	US 1998-95372	19980610
US 2002132348	A1	20020919	US 2000-729043	20001204
PRAI US 1995-8250P	P	19951206		
US 1996-761704	A2	19961206		
US 1996-32645P	P	19961210		
US 1998-95372	A1	19980610		

AB Claimed is a yeast-bacteria shuttle vector comprising both yeast and bacterial replication origins and selection marker genes, where the bacterial replication origin is either from the \*\*\*P1\*\*\* replicon\*\*\* or the \*\*\*F\*\*\* \*\*\*factor\*\*\* origin of replication. The functional anal. of genes frequently requires the manipulation of large genomic regions. A yeast-bacteria shuttle vector is described, that can be used to clone large regions of DNA by homologous recombination. The important feature of present invention is the presence of the a bacterial replication origin, which allows large DNA insert capacity. The utility of this vector lies in its ability to isolate, manipulate and maintain large fragments in bacteria and yeast, allowing for mutagenesis by yeast genetics and simplified prep. of plasmid DNA in bacteria.

RE.CNT 17 THERE ARE 17 CITED REFERENCES AVAILABLE FOR THIS RECORD

ALL CITATIONS AVAILABLE IN THE RE FORMAT

=> s p1 and replicon  
L2 250 P1 AND REPLICON

=> s p1 (3a) replicon  
L3 78 P1 (3A) REPLICON

=> s F (3a) factor?  
L4 10056 F (3A) FACTOR?

=> s l3 and l4  
L5 3 L3 AND L4

=> dup rem l5  
PROCESSING COMPLETED FOR L5  
L6 2 DUP REM L5 (1 DUPLICATE REMOVED)

=> s l6 not l1  
L7 1 L6 NOT L1

=> d bib abs

L7 ANSWER 1 OF 1 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS

INC.

AN 1984:298540 BIOSIS

DN BA78:35020

TI P-1 PLASMID REPLICATION REPLICON STRUCTURE.

AU ABELES A L; SNYDER K M; CHATTORAJ D K

CS LAB. MOL. BIOL., LBI-BASIC RES. PROGRAM, NCI-FREDERICK CANCER RES. FAC.,

P.O. BOX B, FREDERICK, MD 21701, USA.

SO J MOL BIOL. (1984) 173 (3), 307-324.

CODEN: JMOBAK. ISSN: 0022-2836.

FS BA; OLD

LA English

AB Phage P1 lysogenizes *Escherichia coli* as a unit-copy plasmid. The plasmid-encoded elements implicated on P1 plasmid maintenance were defined. A 2081 base-pair fragment of the 90,000 base P1 plasmid confers the capacity for controlled plasmid replication. DNA sequence analysis reveals several open reading frames in this fragment. The largest is shown to encode a 32,000 MW protein required for plasmid replication. The corresponding gene, repA, was identified genetically. A set of five 19 base-pair repeats is located upstream from repA; a set of 9 similar repeats is located immediately downstream from repA. Each set of repeats, when cloned into pBR322, exerts incompatibility towards a \*\*\*P1\*\*\*

\*\*\*replicon\*\*\*. The upstream set, designated incC, consists of direct repeats that are spaced about 2 turns of the DNA helix apart; the downstream set, designated incA, consists of 9 repeats arranged 3 in one orientation and 6 in the other. Spacing between incA repeats were 3 or 4 turns of the helix apart. The organization of the plasmid maintenance regions of P1 and the unit-copy sex \*\*\*factor\*\*\* plasmid, \*\*\*F\*\*\*, is strikingly similar. Although the DNA sequences of this region in the 2 plasmids exhibit little homology, a 9 base-pair sequence that appears 4 times in the origin region of members of the Enterobacteriaceae also occurs twice as direct repeats in similar positions in P1 and F. This sequence, where it occurs in *E. coli*, was postulated to be the binding site for the essential replication protein determined by dnaA. The dnaA protein appears not to be essential for the replication of either plasmid; the function of the sequence in P1 and F may be regulatory.

=> s knock in or knock-out

L8 11698 KNOCK IN OR KNOCK-OUT

=> d his

(FILE 'HOME' ENTERED AT 12:04:34 ON 11 DEC 2002)

FILE 'BIOSIS, EMBASE, CAPLUS' ENTERED AT 12:04:41 ON 11 DEC 2002

L1 1 S P1 REPLICON AND F FACTOR

L2 250 S P1 AND REPLICON

L3 78 S P1 (3A) REPLICON

L4 10056 S F (3A) FACTOR?

L5 3 S L3 AND L4

L6 2 DUP REM L5 (1 DUPLICATE REMOVED)

L7 1 S L6 NOT L1

L8 11698 S KNOCK IN OR KNOCK-OUT

=> s l3 and l8

L9 0 L3 AND L8

=> s homologo? recomb? (3s) yeast

L10 1498 HOMOLOGO? RECOMBIN? (3S) YEAST

=> s l10 and l8

L11 1 L10 AND L8

=> dup rem l11

PROCESSING COMPLETED FOR L11

L12 8 DUP REM L11 (6 DUPLICATES REMOVED)

=> d bib abs 1-

YOU HAVE REQUESTED DATA FROM 8 ANSWERS - CONTINUE? Y/(N):y

L12 ANSWER 1 OF 8 CAPLUS COPYRIGHT 2002 ACS

AN 2002:794235 CAPLUS

DN 137:274176

TI Homologous recombination in mismatch repair inactivated eukaryotic cells

IN Te Riele, Henricus Petrus Joseph; De Wind, Niels; Dekker-Vlaar, Helena

Maria Johanna

PA Neth.

SO U.S. Pat. Appl. Publ., 18 pp., Cont.-in-part of U.S. Ser. No. 147,712,

abandoned.

CODEN: USXXCO

DT Patent

LA English

FAN.CNT 2

PATENT NO. KIND DATE APPLICATION NO. DATE

PI US 2002151059 A1 20021017 US 2001-884877 20010620

WO 9705268 A1 19970213 WO 1995-EP2980 19950726

W: AU, BR, CA, CN, JP, KR, MX, NO, NZ, RU, SE, SG, US

RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE,

BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG

PRAI WO 1995-EP2980 W 19950726

US 1999-147712 B2 19990223

AB A mammalian cell having a mismatch repair-deficient phenotype is provided, where one or both alleles of a gene essential for mismatch repair, such as an Msh gene, are inactivated. Using this cell in a gene \*\*\*knock\*\*\*

\*\*\*out\*\*\* methodol. advantageously allows efficient homologous recombination, even when the DNA sequences of the donor and recipient sequences diverge by significantly more than 0.6%. The present invention relates to a method for modifying the genome of eukaryotic cells by homologous recombination using DNA sequences which substantially differ from the target locus with respect to the nucleotide sequence (0.1 to 30 % divergence) in the region where recombination takes place. Homologous recombination between diverged sequences is achieved by genetic or transitory inactivation of the cell's mismatch repair system.

L12 ANSWER 2 OF 8 CAPLUS COPYRIGHT 2002 ACS

AN 2001:90488 CAPLUS

DN 136:51264

TI Altering eIF-2.alpha. translation activity in plants in response to environmental stress

IN Roth, Donald A.

PA University of Wyoming, USA

SO PCT Int. Appl., 67 pp.

CODEN: PIXXD2

DT Patent

LA English

FAN.CNT 1

PATENT NO. KIND DATE APPLICATION NO. DATE

PI WO 2001094558 A1 20011213 WO 2001-US18342 20010606

W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MY, NZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, BG, KG, KZ, MD, RU, TJ, TM

RW: GH, GM, KE, LS, MV, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG

PRAI US 2000-587789 A 20000606

AB The present invention relates to compns. and methods useful to alter translation of the eukaryotic initiation factor-2.alpha. (eIF-2.alpha.) in response to biotic and abiotic stress, or growth pattern adjustment in plants. Stress factors can include pathogen attack, wounding, drought, hypoxia, light, or temp. Furthermore, a \*\*\*knock\*\*\* - \*\*\*out\*\*\* construct of the eIF-2.alpha. gene is provided, capable of \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* with the wild-type gene. Wheat eIF2.alpha. was shown to be able to substitute for the \*\*\*yeast\*\*\* eIF2.alpha. in \*\*\*yeast\*\*\* SUI2 mutants. Similarly, it was shown to be functional in mammalian cells where it was phosphorylated by PKR kinase.

RE.CNT 2 THERE ARE 2 CITED REFERENCES AVAILABLE FOR THIS

RECORD

ALL CITATIONS AVAILABLE IN THE RE FORMAT

L12 ANSWER 3 OF 8 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS

INC.DUPLICATE 1

AN 2001:245966 BIOSIS

DN PREV200100245966

TI Efficient rejoining of radiation-induced DNA double-strand breaks in

vertebrate cells deficient in genes of the RAD52 epistasis group.

AU Wang, Huichen; Zeng, Zhao-Chong; Bui, Tu-Anh; Sonoda, Eiichi; Takata,

Minoru; Takeda, Shunichi; Iliakis, George (1)

CS (1) Department of Radiation Oncology of Kimmel Cancer Center, Jefferson

Medical College, Thompson Building Room B-1, Philadelphia, PA, 19107 USA

SO Oncogene, (26 April, 2001) Vol. 20, No. 18, pp. 2212-2224. print

ISSN: 0950-9232.

DT Article

LA English

SL English

AB Rejoining of ionizing radiation (IR) induced DNA DSBs usually follows biphasic kinetics with a fast (t50: 5-30 min) component attributed to DNA-PK-dependent non-homologous endjoining (NHEJ) and a slow (t50: 1-20 h), as of yet uncharacterized, component. To examine whether \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* (HR) contributes to DNA DSB rejoining, a systematic genetic study was undertaken using the hyper-recombinogenic DT40 chicken cell line and a series of mutants defective in HR. We show that DT40 cells rejoin IR-induced DNA DSBs with half times of 13 min and 4.5 h and contributions by the fast (78%) and the slow (22%) components similar to those of other vertebrate cells with 1000-fold lower levels of HR. We also show that deletion of RAD51B, RAD52 and RAD54 leaves unchanged the rejoining half times and the contribution of the slow component, as does also a conditional \*\*\*knock\*\*\* \*\*\*out\*\*\* mutant of RAD51. A significant reduction (to 37%) in the contribution of the fast component is observed in Ku70-/- DT40 cells, but the slow component, operating with a half time of 18.4 h, is still able to rejoin the majority (63%) of DSBs. A double mutant Ku70-/-RAD54-/- shows similar half times to Ku70-/- cells. Thus, variations in HR by several orders of magnitude leave unchanged the kinetics of rejoining of DNA DSBs, and fail to modify the contribution of the slow component in a way compatible with a dependence on HR. We propose that, in contrast to \*\*\*yeast\*\*\*, cells of vertebrates are 'hard-wired' in the utilization of NHEJ as the main pathway for rejoining of IR-induced DNA DSBs and speculate that the contribution of \*\*\*homologous\*\*\*

\*\*\*recombination\*\*\* repair (HRR) is at a stage after the initial rejoining.

L12 ANSWER 4 OF 8 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.

AN 2000:190845 BIOSIS

DN PREV20000190845

TI Bryophytes as model systems.

AU Wood, Andrew J. (1); Oliver, Melvin J.; Cove, David J.

CS (1) Department of Plant Biology, Southern Illinois University-Carbondale, Carbondale, IL, 62901-6509 USA

SO Bryologist, (Spring, 2000) Vol. 103, No. 1, pp. 128-133.

ISSN: 0007-2745.

DT Article

LA English

SL English

AB Bryophytes have been powerful experimental tools for the elucidation of complex biological processes. Analysis of organisms from these ancient clades is an active and ongoing enterprise that will provide greater insight into the development, physiology, phylogenetics, and stress-induced cellular responses of plants. To maintain their relevance as experimental models, the analysis of mosses must expand to include modern molecular tools such as a knowledge of the genome via large-scale DNA sequencing, the ability to create transgenic individuals via transformation, and the capability to create gene \*\*\*knock\*\*\*-outs by \*\*\*homologous\*\*\* \*\*\*recombination\*\*\*. The availability of these molecular tools is limited when compared to flowering plants. However, in mosses such as *Physcomitrella patens*, *Funaria hygrometrica*, *Ceratodon purpureus*, and *Tortula ruralis* these tools are rapidly being developed for the study of molecular genetics. Efficient targeted gene disruption (i.e., \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* ) is a well-established tool in both \*\*\*yeast\*\*\* and murine cells that until recently was unknown in any plant model system. Recently, Schaefer and Zryd (1997) demonstrated that efficient \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* occurs in *P. patens*. The ability to perform efficient \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* in *P. patens* is at present unique amongst all plants and represents an extremely powerful technique for the functional analysis of plant genes.

L12 ANSWER 5 OF 8 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 2

AN 1998:165336 BIOSIS

DN PREV199800165336

TI I-SceI-induced gene replacement at a natural locus in embryonic stem cells.

AU Cohen-Tannoudji, Michel; Robine, Sylvie; Choulika, Andre; Pinto, Daniel; El Marjoui, Fatima; Babinet, Charles; Louvard, Daniel; Jaisser, Frederic (1)

CS (1) INSERM U246, Faculte de Medecine X. Bichat, 16 rue H. Huchard, 75018 Paris France

SO Molecular and Cellular Biology, (March, 1998) Vol. 18, No. 3, pp. 1444-1448.

ISSN: 0270-7306.

DT Article

LA English

AB Gene targeting is a very powerful tool for studying mammalian development and physiology and for creating models of human diseases. In many instances, however, it is desirable to study different modifications of a target gene, but this is limited by the generally low frequency of \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* in mammalian cells. We have developed a novel gene-targeting strategy in mouse embryonic stem cells that is based on the induction of endogenous gap repair processes at a defined location within the genome by induction of a double-strand break (DSB) in the gene to be mutated. This strategy was used to \*\*\*knock\*\*\* in an NH2-ezrin mutant in the villin gene, which encodes an actin-binding protein expressed in the brush border of the intestine and the kidney. To induce the DSB, an I-SceI \*\*\*yeast\*\*\* meganuclease restriction site was first introduced by gene targeting to the villin gene, followed by transient expression of I-SceI. The repair of the ensuing DSB was achieved with high efficiency (6 X 10<sup>-6</sup>) by a repair shuttle vector sharing only a 2.8-kb region of homology with the villin gene and no negative selection marker. Compared to conventional gene-targeting experiments at the villin locus, this represents a 100-fold stimulation of gene-targeting frequency, notwithstanding a much lower length of homology. This strategy will be very helpful in facilitating the targeted introduction of several types of mutations within a gene of interest.

L12 ANSWER 6 OF 8 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 3

AN 1997:345271 BIOSIS

DN PREV19979964474

TI Efficient gene targeting in the moss *Physcomitrella patens*.

AU Schaefer, Didier G. (1); Zryd, Jean-Pierre

CS (1) Laboratoire de Phyto-genetique Cellulaire, Universite de Lausanne, Batiment de Biologie, CH-1015 Lausanne-Dorigny Switzerland

SO Plant Journal, (1997) Vol. 11, No. 6, pp. 1195-1206.

ISSN: 0960-7412.

DT Article

LA English

AB The moss *Physcomitrella patens* is used as a genetic model system to study plant development, taking advantage of the fact that the haploid gametophyte dominates in its life cycle. Transformation experiments designed to target three single-copy genomic loci were performed to

determine the efficiency of gene targeting in this plant. Mean transformation rates were 10-fold higher with the targeting vectors and molecular evidence for the integration of exogenous DNA into each targeted locus by \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* is provided. The efficiency of gene targeting determined in these experiments is above 90%, which is in the range of that observed in \*\*\*yeast\*\*\* and several orders of magnitude higher than previous reports of gene targeting in plants. Thus, gene \*\*\*knock\*\*\* - \*\*\*out\*\*\* and allele replacement approaches are directly accessible to study plant development in the moss *Physcomitrella patens*. Moreover, efficient gene targeting has so far only been observed in lower eukaryotes such as protozoa, yeasts and filamentous fungi, and, as shown here the first example from the plant kingdom is a haplobiontic moss. This suggests a possible correlation between efficient gene targeting and haplophase in eukaryotes.

L12 ANSWER 7 OF 8 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 4

AN 1997:391337 BIOSIS

DN PREV199799690540

TI Transposon-generated ' \*\*\*knock\*\*\* - \*\*\*out\*\*\* ' and ' \*\*\*knock\*\*\* -in' gene-targeting constructs for use in mice.

AU Westphal, Christoph Heiner (1); Leder, Philip

CS (1) Dep. Genetics, Harv. Med. Sch., 200 Longwood Ave., Boston, MA 02115 USA

SO Current Biology, (1997) Vol. 7, No. 7, pp. 530-533.

ISSN: 0960-9822.

DT Article

LA English

AB The conventional technique for targeted mutation of mouse genes entails placing a genomic DNA fragment containing the gene of interest into a vector for fine mapping, followed by cloning of two genomic arms around a selectable neomycin-resistance cassette in a vector containing thymidine kinase (1); this generally requires 1-2 months of work for each construct. The single ' \*\*\*knock\*\*\* - \*\*\*out\*\*\* ' construct is then transfected into mouse embryonic stem (ES) cells, which are subsequently subjected to positive selection (using G418 to select for neomycin-resistance) and negative selection (using FIAU to exclude cells lacking thymidine kinase), allowing the selection of cells which have undergone \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* with the knock-out vector. This approach leads to inactivation of the gene of interest (2). Recently, an in vitro reaction was developed, on the basis of the \*\*\*yeast\*\*\* Ty transposon, as a useful technique in shotgun sequencing (3). An artificial transposable element, integrase enzyme and the target plasmid are incubated together to engender transposition. The DNA is then purified, and subsequently electroporated into bacteria. The transposon and the target plasmid bear distinct antibiotic resistance markers (trimethoprim and ampicillin, respectively), allowing double selection for transposition events. In the present study, we have modified this system to allow the rapid, simultaneous generation of a palette of potential gene targeting constructs. Our approach led from genomic clone to completed construct ready for transfection in a matter of days. The results presented here indicate that this technique should also be applicable to the generation of gene fusion constructs (4-8), simplifying this technically demanding method.

L12 ANSWER 8 OF 8 EMBASE COPYRIGHT 2002 ELSEVIER SCI. B.V.

AN 97000851 EMBASE

DN 1997000851

TI Recombinant DNA technology as an investigative tool in drug metabolism research.

AU Friedberg T.; Wolf C.R.

CS T. Friedberg, Biomedical Research Centre, Ninewells Hospital/Medical School, Dundee DD1 9SY, United Kingdom

SO Advanced Drug Delivery Reviews, (1996) 22/1-2 (187-213).

ISSN: 0169-409X CODEN: ADDREP

PUI S 0169-409X(96)00419-X

CY Netherlands

DT Journal; General Review

FS 022 Human Genetics

029 Clinical Biochemistry

036 Health Policy, Economics and Management

052 Toxicology

030 Pharmacology

037 Drug Literature Index

LA English

SL English

AB Drug metabolism influences the pharmacological properties of a vast array of compounds and is controlled by a complex system of drug metabolizing enzymes. A thorough understanding of this system allows the more effective development of therapeutic drugs, as well as a significant improvement of risk assessment, particularly in the field of chemical carcinogenesis. The early identification of potential therapeutic problems relating to drug metabolism could reduce the development costs for pharmaceuticals. Recently, techniques using recombinant DNA have become available for this purpose. In these approaches the genetic information for the enzyme under investigation is expressed in vitro or in vivo, following gene transfer. This approach is called heterologous expression. In addition it is possible to inactivate genes in cells and animals by \*\*\*homologous\*\*\* \*\*\*recombination\*\*\* (gene-targeting, - \*\*\*knock\*\*\* - \*\*\*out\*\*\* ). Heterologous expression and gene \*\*\*knock\*\*\* -outs can be used to define the catalytic parameters as well as the biological role of xenobiotic metabolizing enzymes. Some heterologous expression systems supply sufficient amounts of these enzymes for structure/function

analysis, thus immensely improving the prospects of rational drug design. In addition, these systems provide the basis for rapidly generating immunological tools for the selective quantitation of xenobiotic metabolizing enzymes in human tissues. This combined with the knowledge about the catalytic parameters of a particular enzyme, allows predictions on the exact role of enzymes in drug metabolism as well as drug-drug interactions to be made. However in this regard an important and unfortunately often neglected issue is the appropriate validation of the different heterologous expression systems. Transgenes have also been used to study the regulation of drug metabolizing enzymes by endogenous and exogenous substances using reporter constructs. These studies may also lead to a thorough understanding of the mechanisms underlying interindividual differences in the level of xenobiotic metabolizing enzymes. This article surveys and critically examines the applicability of the different mammalian, \*\*\*yeast\*\*\*, insect and bacterial systems for evaluating the structure, the enzymatic function, the biological role and the regulation of drug metabolizing enzymes in vitro and in vivo.

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---Logging off of STN---

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Executing the logoff script...

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